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# Binding of STW 5 (Iberogast<sup> $\mathbb{R}$ </sup>) and its components to intestinal 5-HT, muscarinic M<sub>3</sub>, and opioid receptors

U. Simmen<sup>a</sup>, O. Kelber<sup>b,\*</sup>, S.N. Okpanyi<sup>b</sup>, R. Jaeggi<sup>c</sup>, B. Bueter<sup>c</sup>, D. Weiser<sup>b</sup>

<sup>a</sup>Institute of Pharmaceutical Biology, University of Basel, Switzerland <sup>b</sup>Steigerwald Arzneimittelwerk GmbH, Darmstadt, Germany

<sup>c</sup>Vitaplant AG, Witterswil, Switzerland

## Abstract

Clinical studies with the fixed herbal combination product STW 5 (Iberogast<sup>®</sup>) have indicated an efficacy comparable to metoclopramide (5-HT<sub>3</sub> antagonist) and cisapride (5-HT<sub>4</sub> agonist) in functional gastro-intestinal diseases like functional dyspepsia (FD) and irritable bowel syndrome (IBS). Since serotonin  $(5-HT_3 \text{ and } 5-HT_4)$  and muscarinic M<sub>3</sub> receptors are known to play a central role in the etiology of FD and IBS, the extracts contained in STW 5 and several of their phytochemical components were studied in vitro for binding affinities to these receptors of the intestine. STW 5 inhibited the binding of <sup>3</sup>H-GR113808 and <sup>3</sup>H-4-DAMP to 5-HT<sub>4</sub> and M<sub>3</sub> receptors, respectively, about 10 times more potently than the binding of <sup>3</sup>H-GR65630 to 5-HT<sub>3</sub> receptors. IC<sub>50</sub> values for STW 5 did correspond to extract dilutions of 1:1000 (M<sub>3</sub> binding) and 1:2000 (5-HT<sub>4</sub> binding). In addition, STW 5 also potently inhibited the binding to opioid receptors with an  $IC_{50}$  value of 1:2000. Of the nine herbal extracts contained in STW 5, the fresh plant extract of bitter candy tuft (*Iberis amara*) selectively inhibited binding to M<sub>3</sub> receptors, while ethanolic extracts of celandine herb and chamomile flower were selective to 5-HT<sub>4</sub>, and liquorice root to 5-HT<sub>3</sub> receptors. Binding affinities to human recombinant 5-HT<sub>3</sub>, 5-HT<sub>4</sub> and M<sub>3</sub> receptors were qualitatively similar to those of the corresponding intestinal receptors. The benzylisoquinoline alkaloid berberine had significant inhibitory action on 5-HT<sub>4</sub> and M<sub>3</sub> binding, showing IC<sub>50</sub> values of 40 ng/ml (100 nM) and 200 ng/ml (500 nM), respectively, but is present in the extract of celandine herb only in traces, so that also for the celandine extract a cooperative effect of several phytochemical constituents can be assumed. These in vitro data indicate that 5-HT<sub>4</sub> (to a lesser degree 5-HT<sub>3</sub>), muscarinic  $M_3$ , and opioid receptors represent target sites for the treatment of FD and IBS with STW 5 (Iberogast<sup>®</sup>). © 2006 Elsevier GmbH. All rights reserved.

*Keywords:* Receptor binding; Serotonin 5-HT<sub>3</sub>; 5-HT<sub>4</sub>; Muscarinic M<sub>3</sub>; Opioid receptors; Functional dyspepsia (FD); Irritable bowel syndrome (IBS); STW 5; Iberogast; *Iberis amara; Chamomila recutita; Mentha piperita; Carum carvi; Chelidonium majus; Silybum marianum; Angelica archangelica; Glycyrrhiza glabra; Melissa officinalis* 

## Introduction

\*Corresponding author. Tel.: +4961513305154; fax: +4961513305493.

*E-mail address:* kelber@steigerwald.de (O. Kelber).

The fixed herbal product STW 5 (Iberogast<sup>®</sup>) is composed of 15 ml of a fresh plant extract (1:2, 50% v/v ethanol) of bitter candy tuft and extracts (1:3, 30% v/v ethanol) of angelica root (10 ml), milk thistle fruit

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(10 ml), caraway fruit (10 ml), celandine herb (10 ml), liquorice root (10 ml), chamomile flower (20 ml), lemon balm leaf (10 ml) and peppermint leaf (5 ml) per 100 ml. It has been clinically proven to be effective against nonulcer dyspepsia and irritable bowel syndrome (Saller et al., 2002; Rösch et al. 2006) with similar efficacy but less side effects as cisapride (Roesch et al., 2002) and metoclopramide (Nikolay, 1984).

In pharmacological studies with the isolated guinea pig ileum, stimulated with acetvlcholine or histamine, STW 5 has been shown to exhibit spasmolytic activities as well as the extracts of chamomile flowers. liquorice root, and peppermint leaves (Ammon et al., 2006). In the non-stimulated ileum preparation STW 5 and its component Iberis amara raised the basal tone of the smooth muscles (Okpanyi et al., 1993; Ammon et al., 2006; Michael et al., 2006). In the stomach, a region specific effect was described in vitro (Schemann et al., 2006), which has been confirmed in a clinical pharmacological study, too (Pilichiewicz et al., 2006). Besides these motility-related effects an effect on gastro-intestinal sensitivity (Müller et al., 2006) and a dose dependent anti-ulcerogenic activity as well as cytoprotective effects were shown in in vivo studies (Khayyal et al., 2006). The uptake of characteristic constituents of the extracts by the small intestine has been proven in an ex vivo model (Kelber et al., 2006).

Functional gastro-intestinal diseases like functional dyspepsia (FD) and irritable bowel syndrome (IBS) are some of the most common ailments for medical treatment in industrialized countries (Allescher, 2006). Synthetic drugs like alosetron  $(5-HT_3 \text{ antagonist})$ , cisapride and tegaserod (both 5-HT<sub>4</sub> agonists) have recently been developed. But despite promising pharmacological evidence, their clinical use remains restricted: Alosetron and cisapride have been removed from the market, because of severe adverse effects ischemic colitis and cardiac like arrhythmias (Callahan, 2002; Kamm, 2002). However, cisapride has recently been reapproved under restrictions in certain countries. Whether the partial agonist tegaserod can prevail as clinical remedy, remains to be seen. Nevertheless, there is significant pharmacological evidence that 5-HT<sub>3</sub> and 5-HT<sub>4</sub> receptors of the submucosal sensory neurons as well as the muscarinic M<sub>3</sub> receptor of smooth muscle cells represent promising targets for the treatment of functional gastro-intestinal diseases (Houghton et al., 1997; Prins et al., 1999; Berman et al., 2002). Moreover, opioid receptor agonists have also been discussed as new drugs in the treatment of FD (Stanghellini et al., 2003; Camilleri 2004). Since the effect of STW 5 on these intestinal receptors had not been studied yet, competitive binding experiments with STW 5 components and its characteristic constituents were performed.

#### Materials and methods

Ethanolic extracts (30% v/v ethanol) of STW 5 were diluted with H<sub>2</sub>O to prepare the following extract dilutions studied in the binding experiments: 1:200, 1:500, 1:1000, and 1:10,000. Cisapride was purchased from RDI (Flanders, NJ, USA). Intestine of the rat (Wistar, RCC Ltd., Switzerland) was homogenized in 50 mM Tris/HCl pH 7.4 on ice using a Polytron homogenizer. The homogenate was centrifuged at 38,000*g* for 20 min and washed five times. The preparation was stored as a pellet at  $-80 \,^{\circ}$ C and was homogenized each time when used for binding studies. The protein content of the intestine for each binding assay was about 20 µg/ml as determined by the BCA protein assay.

Binding assays to 5-HT<sub>3</sub>, 5-HT<sub>4</sub>, muscarinic M<sub>3</sub>, 5-HT<sub>1A</sub>, and opioid receptors of the intestine were performed with <sup>3</sup>H-GR65630 (5-HT<sub>3</sub> antagonist, 0.2 nM), <sup>3</sup>H-GR113808 (5-HT<sub>4</sub> antagonist, 0.2 nM), (N-methyl-<sup>3</sup>H) 4-DAMP ( $M_3$  antagonist, 0.2 nM), <sup>3</sup>H-8-OH-DPAT (5-HT<sub>1A</sub> agonist, 0.5 nM), and <sup>3</sup>H-naloxone (non-selective antagonist, 3.6 nM), while non-specific binding was determined in the presence of metoclopramide, 5-HT, atropine, 8-OH-DPAT (each  $10^{-4}$  M), and naloxone (2 ×  $10^{-5}$ ), respectively. Binding equilibration was terminated by filtration (Whatman GF/C filter) and radioactivity was quantified by liquid scintillation counting. IC<sub>50</sub> values expressed as extract dilution were graphically determined according to concentration response curves of the specific binding (difference between specific and non-specific binding in the presence of given extract concentrations).

#### **Results and discussion**

STW 5 and its nine herbal component extracts affected the binding to 5-HT<sub>3</sub>, 5-HT<sub>4</sub> and muscarinic  $M_3$  with different potencies as summarized in Table 1. STW 5 showed a >10 fold higher affinity to both 5-HT<sub>4</sub> and  $M_3$  receptors than to 5-HT<sub>3</sub> receptors. IC<sub>50</sub> values for the fixed combination STW 5 did correspond to dilutions of 1:2000 (5-HT<sub>4</sub> binding) and 1:1000 (M<sub>3</sub> binding). Whether these pharmacological actions on 5-HT<sub>4</sub> and M<sub>3</sub> receptors account for the observed effects in vivo, cannot directly be concluded. Considering a dilution factor of approximately 1:1000 for STW 5 extracts entering the stomach (1 ml drug in 11 gastrointestinal volume), in vivo effects seem to be plausible for seven of the nine component extracts that showed  $IC_{50}$  values <1:1000. Among the nine herbal extracts, the fresh plant extract of bitter candy tuft selectively inhibited binding to M3 receptors, while ethanolic extracts of celandine herb and chamomile flowers were

**Table 1.** Binding affinities of STW 5 and its components to rat intestinal 5-HT<sub>3</sub>, 5-HT<sub>4</sub> and muscarinic M<sub>3</sub> receptors expressed as IC<sub>50</sub> values

	IC <sub>50</sub> (extract dilution)		
	5-HT <sub>3</sub>	5-HT <sub>4</sub>	$M_3$
Iberogast <sup>®</sup> (STW 5)	<1:100	1:2000	1:1000
Peppermint leaf (STW 5-KII)	a	1:2000	b
Chamomile flower (STW 5-KIII)	a	1:10,000	b
Liquorice root (STW 5-KIV)	1:2000	1:500	b
Angelica root (STW 5-KV)	1:2000	1:2000	1:1250
Caraway fruit (STW 5-KVI)	a	a	b
Milk thistle fruit (STW 5-KVII)	a	a	b
Melissa leaf (STW 5-KVIII)	<1:100	1:1000	b
Celandine herb (STW 5-KIX)	1:1000	>1:10,000	1:3500
Bitter candy tuft (STW 6)	<1:100	1:500	1:1500

 $^{a}$ High non-specific binding in the presence of extract unables determination of IC<sub>50</sub> value.

 $^{\rm b}$ Specific binding was inhibited <20% in the presence of extract dilutions of 1:2000.

**Table 2.** Binding affinities of STW 5 and components to human recombinant 5-HT<sub>3</sub>, 5-HT<sub>4</sub> and muscarinic M<sub>3</sub> receptors expressed as IC<sub>50</sub> values

	IC <sub>50</sub> (extract dilution)		
	5-HT <sub>3</sub>	5-HT4	$M_3$
Iberogast <sup>®</sup> (STW 5) Liquorice root (STW 5-KIV) Celandine herb (STW 5-KIX) Bitter candy tuft (STW 6)	1:50 1:10 n.d. n.d.	1:160 n.d. 1:330 n.d.	1:125 n.d. n.d. <1:100

selective to 5-HT<sub>4</sub>, and liquorice roots to 5-HT<sub>3</sub> receptors. For caraway and milk thistle fruit extracts, no receptor affinity could be detected. These two extracts exhibited high non-specific binding making an  $IC_{50}$  determination impossible. Such interfering effects can usually be prevented by elimination of the corresponding compounds via extract fractionation. It is therefore possible that parts of these extracts lacking these interfering constituents may also show binding affinity to the receptors studied.

STW 5 and selected component extracts showed qualitatively similar binding affinities to human recombinant 5-HT<sub>3</sub>, 5-HT<sub>4</sub> and muscarinic M<sub>3</sub> receptors as to the corresponding intestinal receptors (Table 2). The binding affinities of STW 5, extracts of liquorice roots, celandine herb and *Iberis amara* to the recombinant receptors were, however, approximately 10 times lower than to the intestinal receptors, probably due to a lower specific binding of the tritiated ligands.

Specific binding of STW 5 and its component extracts chamomile flowers and celandine herb to 5-HT<sub>4</sub>

100 90 80 70 Specific Binding (%) 60 50 40 30 20 10 0 -10 0.00001 0.0001 0.001 0.01 0.1 Extract dilution

**Fig. 1.** Competition for <sup>3</sup>H-GR113808 binding to intestinal 5-HT<sub>4</sub> receptors in the presence of STW 5 ( $\blacksquare$ ), chamomile flower ( $\bigcirc$ ) and celandine herb ( $\triangle$ ). Data are given as mean values  $\pm$  S.D. of three replicates.

**Table 3.** Binding affinities of STW 5 to rat intestinal 5-HT<sub>1A</sub> and opioid receptors expressed as IC<sub>50</sub> values

	IC <sub>50</sub> (extract dilution)		
	5-HT <sub>1A</sub>	Opioid (non-selective)	
STW 5 (Iberogast <sup>®</sup> )	a	1:2000	

<sup>a</sup>No inhibition at extract dilutions between 1:2000 and 1:500.

receptors is shown in Fig. 1. Four extract dilutions (1:200, 1:500, 1:2000, and 1:10,000) were tested at each case to graphically determine  $IC_{50}$  values. Both extracts evidently inhibited the binding of <sup>3</sup>H-GR113808 more potently than the other components of STW 5 indicating their relevance in the fixed herbal combination.

STW 5 also potently inhibited the non-selective binding of <sup>3</sup>H-naloxone to intestinal opioid receptors with an IC<sub>50</sub> value of 1: 2000 (Table 3). No binding inhibition up to a dilution of 1:500 was observed to the intestinal 5-HT<sub>1A</sub> receptor.

Binding affinities of characteristic constituents of STW 5 components to 5-HT<sub>4</sub> and M<sub>3</sub> receptors are summarized in Table 4. Beside berberine chloride, none of the constituents tested significantly inhibited <sup>3</sup>H-GR113808 and (*N*-methyl-<sup>3</sup>H) 4-DAMP binding. Berberine chloride inhibited with similar potency as the reference 5-HT<sub>4</sub> agonist cisapride. Whether berberine also exhibits an action in vivo is doubtful, since the benzylisoquinoline alkaloid is present in extracts of celandine herb (STW 5-K IX) only in traces (total

Table 4. Binding affinities of STW 5 constituents to rat intestinal 5-HT<sub>4</sub> and  $M_3$  receptors expressed as  $IC_{50}$  values

	IC <sub>50</sub> (µg/ml)		
	5-KHT <sub>4</sub>	M <sub>3</sub>	
Cucurbitacin E <sup>1</sup>	>20	>10	
Cucurbitacin I <sup>1</sup>	>20	10	
Glucoiberin <sup>1</sup>	>20	>10	
Kaempferol-3,4-β-	>20	>10	
diglucopyranoside-7-			
O-rhamnopyranoside <sup>1</sup>			
α-Bisabolol <sup>2</sup>	>20	n.d.	
Matricine <sup>2</sup>	>20	n.d.	
Chamazulene <sup>2</sup>	>20	n.d.	
<i>N</i> -malonyltryptophan <sup>3</sup>	>20	>10	
Chelidonic acid <sup>4</sup>	>20	>10	
Chelidonine <sup>4</sup>	>20	10	
Berberine chloride <sup>4</sup>	0.04	0.2	
Sparteine <sup>4</sup>	n.d.	10	
Osthole <sup>5</sup>	n.d.	>10	
Angelicine <sup>5</sup>	n.d.	>10	
Bergaptene <sup>5</sup>	n.d.	10	
Psoralene <sup>5</sup>	n.d.	10	
Chlorogenic acid <sup>5</sup>	n.d.	10	
Umbelliferone <sup>5</sup>	n.d.	10	
Angelicalactone <sup>5</sup>	n.d.	10	
Cisapride	0.025		
Atropine	0.07		

Compound contained in extracts of <sup>1</sup>Bitter candy fruit, <sup>2</sup>Camomile flower, <sup>3</sup>Milk thistle fruit, <sup>4</sup>Celandine herb, <sup>5</sup>Angelica root; n.d. not determined.

alkaloid content <0.8 mg/g). Since none of the remaining characteristic constituents of celandine herb (chelidonine and chelidonic acid) and chamomile flowers ( $\alpha$ -bisabolol, matricine and its degradation product chamazulene) inhibited the binding on 5-HT<sub>4</sub> and M<sub>3</sub> receptors, other constituents seem to account for the observed effects of the extracts. The phenomenon that single compounds fail to explain the effect of complex herbal mixtures seems to be a common case in such in vitro test systems. An explanation for this phenomenon may be that weak effects of various constituents interact in an unknown manner (e.g. additive, competitive, synergistic, etc.), which has to be looked further into in future studies.

# Conclusions

Iberogast<sup>®</sup> (STW 5) has been shown to exhibit binding affinity to intestinal 5-HT<sub>3</sub>, 5-HT<sub>4</sub>, muscarinic M<sub>3</sub>, and opioid receptors with a >10 fold preference for the three latter receptor types. The comparison of the nine herbal component extracts present in STW 5 with regard to binding affinity to these receptors revealed a different pattern of selectivity. Whereas a fresh plant extract of *Iberis amara* selectively inhibited binding to  $M_3$  receptors, ethanolic extracts of celandine herb and chamomile flowers were selective to 5-HT<sub>4</sub>, and liquorice roots to 5-HT<sub>3</sub> receptors. These findings indicate the involvement into receptor binding of seven of the nine components of STW 5 used for functional gastro-intestinal diseases like non-ulcer dyspepsia and irritable bowel syndrome. In conclusion, our in vitro data confirm the relevance of the components of STW 5 for its efficacy in the treatment of functional gastro-intestinal diseases acting via crucial target sites including 5-HT<sub>3</sub>, 5-HT<sub>4</sub>, muscarinic M<sub>3</sub>, and opioid receptors.

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